

RESEARCH ARTICLE

Cytoprotective role of Vitamin E on the toxicity of ABVD (doxorubicin, bleomycin, vinblastine, and dacarbazine) and COPP (cyclophosphamide, vincristine, procarbazine, and prednisone) chemotherapeutic regimes – An experimental animal study in albino rats

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ABSTRACT

Background: This experimental animal study was designed to evaluate the toxicity profile of two frontline chemotherapeutic regimens used in Hodgkin's lymphoma – doxorubicin, bleomycin, vinblastine, and dacarbazine (ABVD) and cyclophosphamide, vincristine, procarbazine, and prednisone (COPP), which are regimens commonly prescribed. We conducted this experimental animal study using albino rats, incorporating an antioxidant, Vitamin E, to assess if its cytoprotective properties could counter the free radical inducing actions of the two regimens. **Aims and Objectives:** The main objectives of our study were to document histologically, the toxic effects of chemotherapeutic agents in ABVD and COPP regimens in the various organ systems in albino rats, and to assess the protective role of the antioxidant, Vitamin E on drug-induced toxicities in albino rats. **Materials and Methods:** In this experimental animal study, 30 albino rats weighing 150 g–250 g were selected and grouped as control, chemotherapy receiving group, and cytoprotective group receiving ABVD/COPP and Vitamin E. The duration of the study was 4 weeks and hematological and histopathological studies were conducted on the models. Results: The experimental study revealed hematological and organ toxicities. Cytoprotection by Vitamin E was evident in the liver and stomach specimens on histopathology. Protection appeared minimal against hematological toxicity. **Conclusions:** The experimental animal study demonstrated a cytoprotective effect of Vitamin E against chemotherapy induced organ toxicity mainly in the stomach and liver, of the animals studied.

KEY WORDS: Cytoprotective; Vitamin E; ABVD; COPP; Albino Rats


INTRODUCTION

Our study is an experimental study on albino rats, focusing on the toxicity profile of two chemotherapeutic regimens

employed in Hodgkin's lymphoma – doxorubicin, bleomycin, vinblastine, and dacarbazine (ABVD) and cyclophosphamide, vincristine, procarbazine, and prednisone (COPP), which are commonly prescribed frontline regimens.^[1,2] Furthermore, we conducted an experimental animal study incorporating Vitamin E, an antioxidant which has received great attention the world over; to assess if its cytoprotective properties could counter the free radical inducing actions of the two regimens.

Objectives

The objectives of the study were as follows:

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- To document histologically, the toxic effects of chemotherapeutic agents in ABVD and COPP regimens in the various organ systems in albino rats
- To assess the protective role of the antioxidant, Vitamin E against these toxicities in albino rats.

MATERIALS AND METHODS

The experimental study involved 30 albino rats (*Rattus norvegicus*) maintained under identical conditions in the animal house; weighing about 150 g–250 g. The Institutional Animal Ethics Committee approval was sought before conducting the study and all experiments were conducted following the guidelines set by the Institutional Animal Ethics Committee. The animals were divided basically into three groups; Groups I, II, and III; for convenience of statistical analysis. Groups II and III were again subdivided into two [Table 1]. Hence, overall, there were five groups, with six rats in each group; maintaining the group average weight equal.

The five groups were:

1. Group I: Control
2. Group II: Anticancer drugs alone
 - II a – receiving ABVD
 - II b – receiving COPP.
3. Group III: Cytoprotective group
 - III a – receiving ABVD and Vitamin E
 - III b – receiving COPP and Vitamin E.

The duration of the study was 4 weeks. The first group, taken as control, was given distilled water. The second group and third group were given ABVD and COPP, respectively; while the fourth and fifth groups were given anticancer drugs along with Vitamin E, an antioxidant [Table 1]. The physical attributes of the five groups are given in Table 2.

The species, sex, weight, the test substance, time of injection, the dose, solvent, and the route were recorded. The dose and routes used for the administration of drugs are given in Table 3. General signs and changes in behavioral, neurological, and autonomic profile are noted^[2] (Tables 4-6). Hematological investigations such as hemoglobin, total count, platelet count estimations, and measurement of weight were done during and at the end of the study.

General Signs

- Heart rate
- Respiratory rate
- Skin color.

Tail Clip Method^[3,4]

This is a test for detecting neuropathy by estimating reaction time. A bulldog clamp with thin rubber sleeves

Table 1: Study groups of the animal models

| Groups | Subdivisions | n=6 |
|---------------------------|--------------|---------------------------|
| I Control | I | Control (distilled water) |
| II Anticancer drugs alone | II a | ABVD |
| | II b | COPP |
| III Cytoprotective group | III a | ABVD + Vit E |
| | III b | COPP + Vit E |

ABVD: Doxorubicin, bleomycin, vinblastine, and dacarbazine,
COPP: Cyclophosphamide, vincristine, procarbazine, and prednisone

is applied to the base of the rat's tail for 30 s. Control rats make continuous efforts to dislodge the clip by biting it. This is taken as the reaction time. Drug-induced neuropathy makes the affected rats indifferent to the clip.

Animals were sacrificed at the end and histopathological examinations of internal organs were performed. The information and different data obtained were recorded in the pro forma sheets.

Statistics

Data were entered into statistical package Microsoft Excel and checked for data entry errors. For normally distributed continuous variables, mean, standard deviation, and analysis of variance (ANOVA) were done. For data not normally distributed, a non-parametric test such as Kruskal–Wallis was carried out. The level of significance was fixed at 5%.

RESULTS

For the purposes of statistical analysis and discussion, Groups IIa and IIIa are designated as ABVD group, and Groups IIb and IIIb are designated COPP group [Table 7].

Hematological

Hemogram: Normal values in rat (*Rattus norvegicus*)

Hb: 14.8 g% (12–17.5 g%)

TC: 14,000 cells/cu mm (5000–25,000 cells/cu mm)

Platelet count: 1,240,000 lacs/cu mm (11–13 lacs/cu mm)

By the end of the 3rd week, blood was drawn for hematological tests. Tests included estimation of hemoglobin, total count, and platelet count. The results of the laboratory tests are given in Tables 7.1 and 7.2, 8.1 and 8.2, and 9.1 and 9.2.

Table 2: Physical parameters of the animals under study

| Parameter | Group I | | Group II a | | Group II b | | Group III a | | Group III b | |
|-----------------------|---------|--------|------------|--------|------------|--------|-------------|--------|-------------|--------|
| | Mean | SD | Mean | SD | Mean | SD | Mean | SD | Mean | SD |
| Weight (g) | 186.66 | 27.32 | 186.61 | 39.88 | 186.49 | 41.312 | 186.60 | 40.33 | 186.53 | 40.12 |
| Length (cm) | 17.66 | 4.597 | 21.167 | 1.602 | 21.333 | 1.966 | 20.167 | 1.552 | 20.333 | 1.666 |
| BSA (m ²) | 0.033 | 0.0036 | 0.032 | 0.0046 | 0.032 | 0.0047 | 0.031 | 0.0046 | 0.030 | 0.0045 |

Table 3: Doses and routes of the drugs used¹

| Regimen | Drugs | Dose | Route |
|-------------|------------------|-----------------------|-------|
| ABVD | Adriamycin | 25 mg/m ² | IP |
| | Bleomycin | 10 mg/m ² | IP |
| | Vinblastine | 6 mg/m ² | IP |
| | Doxorubicin | 375 mg/m ² | IP |
| COPP | Cyclophosphamide | 650 mg/m ² | IP |
| | Vincristine | 1.4 mg/m ² | IP |
| | Prednisolone | 40 mg/m ² | PO |
| | Procarbazine | 100 mg/m ² | IP |
| Antioxidant | Vitamin E | 100mg/kg body wt. | PO |

ABVD: Doxorubicin, bleomycin, vinblastine, and dacarbazine, COPP: Cyclophosphamide, vincristine, procarbazine, and prednisone

Table 4: Assessment of behavioral profile of the animal models

| Profile | Behavior | Interpretation |
|----------------|----------------------|--|
| Awareness | Alert/stuporous | CNS stimulation/depression |
| | Virtual placing | Animal response being placed in different positions; normal or subnormal ability (motor incoordination) |
| | Stereotypy | Frequent mechanical repetition of a movement |
| | Passivity | Animal's response to being placed in unaccustomed positions. (tranquilization, central depression, myorelaxation paralysis, or anesthesia) |
| Mood | Grooming | Excessive indicates central excitation, stimulation |
| | Restlessness | Indicates central stimulation, discomfort; visceral changes |
| | Irritability | Extension of restlessness |
| | Fearfulness | Do not endure gentle manipulation |
| Motor activity | Spontaneous activity | Animal placed in bell jar shows moderate degree of inquisitive behavior |
| | Reactivity | Animal is removed from the jar and placed on a table |
| | Touch response | Animal is touched with a pencil or forceps at various parts (indicates presence of anesthetic activity) |
| | Pain response | A small artery clamp is attached to base of the tail (analgesia, sedation, and central depression) |

Table 5: Assessment of neurological profile of the animal models

| Profile | Behavior | Interpretation |
|----------------------|------------------------------|--|
| Central excitation | Startle response | Response of the animal to a loud noise is recorded |
| | Straub response | Degree of elevation of tail |
| | Tremor | If present, is recorded |
| | Convulsion | If present, is recorded |
| Motor incoordination | Body position; limb position | If deviated much from normal; indicates neuromuscular blockade or central disturbance |
| | Staggering gait | Ataxia by test substance |
| | Abnormal gait | Muscle relaxation related to ataxia |
| | Somersault test | Righting reflex |
| Muscle tone | Limb tone | Grasping the forepaw of the animal and noting the resistance to extension of the paw |
| | Body tone and abdominal tone | Noting the muscle tension in comparison with control animal |
| Reflexes | Pinna reflex | Touching the center of the pinna with a fine instrument |
| | Corneal reflex | Touching the cornea with a wisp of cotton |
| | Ipsilateral reflex | Toe pad is compressed with a forceps which causes the animal to flex its leg in a returning movement |

Table 6: Assessment of autonomic profile of the animal models

| Profile | Behavior | Interpretation |
|-----------------|-------------------|--|
| Optical signs | Pupil Size | Compared before and after injection of test substance |
| | Palpebral opening | Compared before and after injection of test substance |
| Secretory signs | Micturition | Muscarinic activity or irritation of urinary tract by test substance or metabolite |
| | Salivation | Muscarinic activity |

Table 7: Study groups of the animal models*

| Groups | Subdivisions | n=6 |
|---------------------------|--------------|---------------------------|
| I | I | Control (distilled water) |
| II Anticancer drugs alone | II a | ABVD |
| | II b | COPP |
| III Cytoprotective group | III a | ABVD + Vit. E |
| | III b | COPP + Vit. E |

*For the purposes of statistical analysis and discussion, Groups IIa and IIIa are designated as ABVD group, and Groups IIb and IIIb are designated COPP group. ABVD: Doxorubicin, bleomycin, vinblastine, and dacarbazine, COPP: Cyclophosphamide, vincristine, procarbazine, and prednisone

Table 7.1: ABVD group: Hemoglobin values at the 3rd week

| ABVD group | Mean (g %) | Variance (g %) | SD (g %) | F/H |
|------------|------------|----------------|----------|---------|
| I | 12.557 | 2.077 | 1.339 | F=0.107 |
| II a | 12.093 | 3.664 | 1.864 | |
| III a | 12.402 | 3.917 | 1.982 | |

Technique used: ANOVA – one-way analysis of variance. P=0.898862; not significant. ABVD: Doxorubicin, bleomycin, vinblastine, and dacarbazine

Table 7.2: COPP group: Hemoglobin values at the 3rd week

| COPP group | Mean (g %) | Variance (g %) | SD (g %) | F/H |
|------------|------------|----------------|----------|---------|
| I | 12.557 | 2.077 | 1.339 | F=0.103 |
| II b | 11.975 | 3.776 | 1.837 | |
| III b | 12.392 | 3.878 | 1.853 | |

Technique used: ANOVA – one-way analysis of variance. P=0.887662; not significant. COPP: Cyclophosphamide, vincristine, procarbazine, and prednisone

Table 8.1: ABVD group: TC values at the 3rd week

| ABVD group | Mean | Variance | SD | F/H |
|------------|--------|--------------------------|--------|---------|
| I | 10.557 | 1.1867 × 10 ⁶ | 1089.7 | H=7.559 |
| II a | 5093 | 7.864 × 10 ⁵ | 924.6 | |
| III a | 8334 | 5.7067 × 10 ⁵ | 755.42 | |

Non-parametric technique: Kruskal–Wallis; one-way analysis of variance. P=0.022834; significant. ABVD: Doxorubicin, bleomycin, vinblastine, and dacarbazine

Table 8.2: COPP group: TC values at the 3rd week

| COPP group | Mean | Variance | SD | F/H |
|------------|--------|--------------------------|--------|---------|
| I | 10.557 | 1.1867 × 10 ⁶ | 1089.7 | H=7.521 |
| II b | 6191.7 | 8.3642 × 10 ⁵ | 914.6 | |
| III b | 8902 | 3.018 × 10 ⁵ | 791.82 | |

Non-parametric technique: Kruskal–Wallis; one-way analysis of variance. P=0.021962; significant. COPP: Cyclophosphamide, vincristine, procarbazine, and prednisone

Table 9.1: ABVD group: PLT count values at the 3rd week

| ABVD group | Mean | Variance | SD | F/H |
|------------|--------------------------|--------------------------|------------|---------|
| I | 1.060 × 10 ⁶ | 6.1854 × 10 ⁸ | 24,508.557 | H=5.719 |
| II a | 1.0155 × 10 ⁶ | 5.8441 × 10 ⁸ | 24,571.642 | |
| III a | 1.0508 × 10 ⁶ | 6.7456 × 10 ⁸ | 23,755.422 | |

Non-parametric technique: Kruskal–Wallis; one-way analysis of variance. P=0.056; not significant. ABVD: Doxorubicin, bleomycin, vinblastine, and dacarbazine

Table 9.2: COPP group: PLT count values at the 3rd week

| COPP group | Mean | Variance | SD | F/H |
|------------|--------------------------|--------------------------|------------|---------|
| I | 1.060 × 10 ⁶ | 6.1854 × 10 ⁸ | 24,508.557 | H=5.689 |
| II b | 1.0375 × 10 ⁶ | 6.3652 × 10 ⁸ | 28,6914.61 | |
| III b | 1.0235 × 10 ⁶ | 6.0182 × 10 ⁸ | 25,631.982 | |

Non-parametric technique: Kruskal–Wallis; one-way analysis of variance. P=0.05616; not significant. COPP: Cyclophosphamide, vincristine, procarbazine, and prednisone

Table 10.1: ABVD group: Weight at the 4th week

| ABVD group | Mean (g) | Variance (g) | SD (g) | F/H |
|------------|----------|--------------|--------|---------|
| I | 29.113 | 466.911 | 21.557 | F=4.596 |
| II a | -10.003 | 441.003 | 20.642 | |
| III a | 3.583 | 598.744 | 24.422 | |

Technique used: ANOVA – one-way analysis of variance. P=0.027751; significant. ABVD: Doxorubicin, bleomycin, vinblastine, and dacarbazine

Table 10.2: COPP group: Weight at the 4th week

| COPP group | Mean (g) | Variance (g) | SD (g) | F/H |
|------------|----------|--------------|--------|---------|
| I | 29.113 | 466.911 | 21.557 | F=4.517 |
| II b | -10.017 | 446.365 | 20.869 | |
| III b | 3.389 | 586.018 | 23.982 | |

Technique used: ANOVA – one-way analysis of variance. P=0.026836; significant. COPP: Cyclophosphamide, vincristine, procarbazine, and prednisone

Hemogram

- The hemoglobin levels did not show much variation in all the groups studied.

Total Count

- The total count values demonstrated fall in both Groups II and III; but the decline in total count was significantly lesser in the cytoprotective group compared to Group II.

Platelet Count

- The platelet count levels did not show much variation in all the groups studied.

Weight

Weight at the 4th week-basal weight (mean difference):

There is weight loss in both groups receiving the drugs; but the loss in weight in the cytoprotective group is significantly of a much smaller magnitude than the weight loss observed in Group II.

General Assessment

Changes cannot be demonstrated in the behavioral, neurological, and autonomic profiles of the animals under study in effective doses of either ABVD or COPP regimens by blind screening methods. Effective dose failed to elicit peripheral analgesia or neuropathy in the rats under study as demonstrated by the tail clip method. There were no significant differences in reaction time in each group.

Histopathological Studies

The following tables (Tables 11–16) show the results of histopathological examination of the internal organs.

HISTOPATHOLOGICAL STUDIES

Stomach [Plate 1]

Table 11: Histopathological changes in the gastric mucosa

| Gastric surface epithelial necrosis | Group |
|-------------------------------------|-------|
| No necrosis (normal mucosa) | I |
| Full thickness | II |
| Partial thickness | III |

- Microscopy of the gastric specimens revealed that a few rats in Group II showed full-thickness denudation of the gastric mucosa [Plate 1.2]. This finding was never found in the cytoprotective group, where it was merely a partial-thickness loss of the mucosa [Table 10; Plate 1.3]. In a few specimens of the cytoprotective group, even normal mucosa was also found.

Liver [Plate 2]

Table 12: Histopathological changes in the liver

| Hepatic architecture | Group |
|-------------------------|-------|
| Normal | I |
| Subcapsular hemorrhage | II |
| Necrosis | II |
| Periportal inflammation | II |

| | |
|-----------------------------------|-----|
| Sinusoidal congestion (extensive) | II |
| Sinusoidal congestion (mild) | III |
| Fatty change (diffuse) | II |
| Fatty change (focal) | III |
| Regeneration | III |

- Regarding microscopy of the hepatic specimens, toxic effects of a milder intensity were observed in the cytoprotective group in comparison to Group II [Plates 2.3A & 2.3B]. Group II specimens showed a more intense and severe manifestation of toxicity [Table 11; Plates 2.2A, B and C].
- The spleen, heart, kidneys, and lungs demonstrated toxic manifestations of similar magnitude and severity in both Groups II and III [Tables 12-16].

Spleen [Plate 3]

Table 13: Histopathological changes in the spleen

| Splenic architecture | Group |
|-----------------------|---------|
| Normal | I |
| Sinusoidal congestion | II, III |
| Hemorrhage | II, III |

Heart [Plate 4]

Table 14: Histopathological changes in the heart

| Cardiac architecture | Group |
|----------------------|---------|
| Normal | I |
| Congestion | II, III |
| Hemorrhage | II, III |
| Pericarditis | II, III |

Kidney [Plate 5]

Table 15: Histopathological changes in the kidney

| Renal architecture | Group |
|------------------------|---------|
| Normal | I |
| Subcapsular hemorrhage | II, III |
| Parenchymal hemorrhage | II, III |
| Tubular necrosis | II, III |

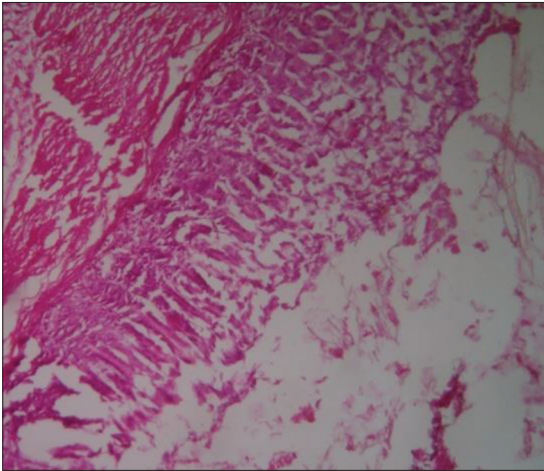
Lung [Plate 6]

Table 16: Histopathological changes in the lung

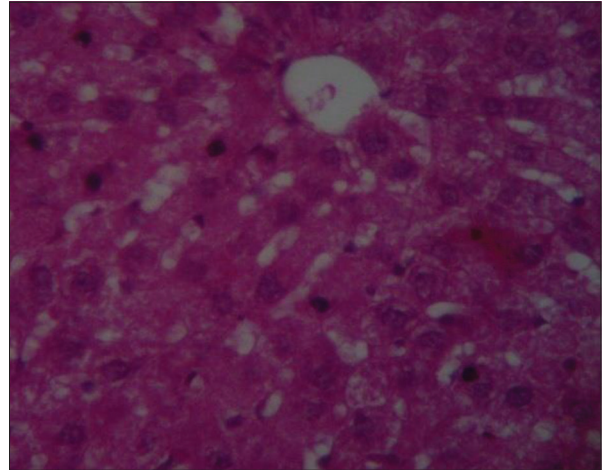
| Pulmonary architecture | Group |
|------------------------|---------|
| Normal | I |
| Inflammation | II, III |
| Collapse | II, III |
| Edema | II, III |

Plate 1 - Experimental animal study: Histopathology of stomach

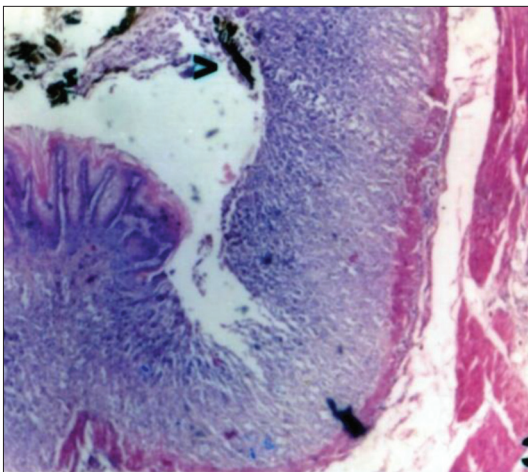
Plate 2 - Experimental animal study: Histopathology of liver



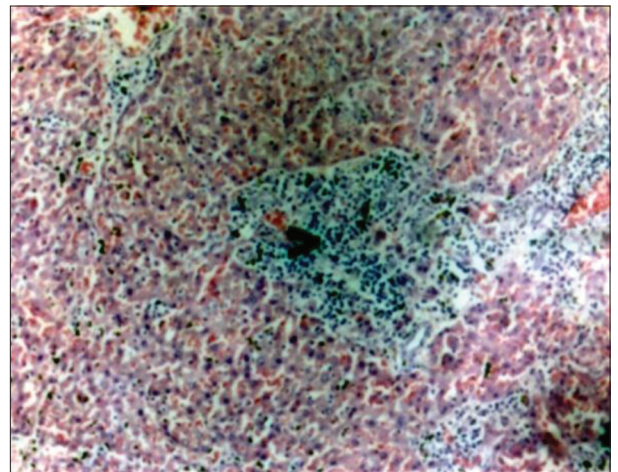
1.1 Group I: Normal gastric mucosa



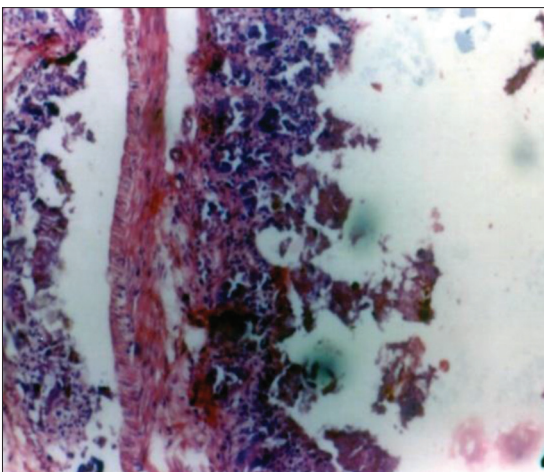
2.1 Group I: Normal liver



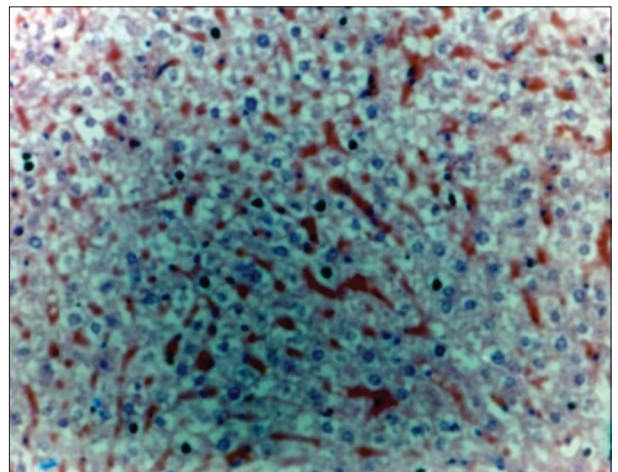
1.2 Group II: Full-thickness mucosal necrosis



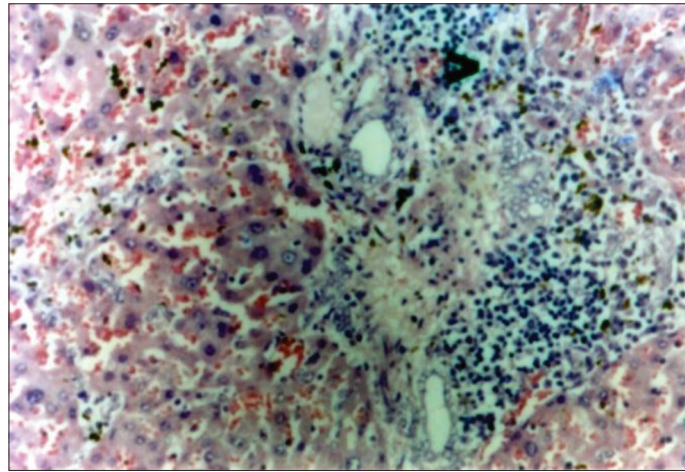
2.2A Group II: Severe congestion and hepatic necrosis



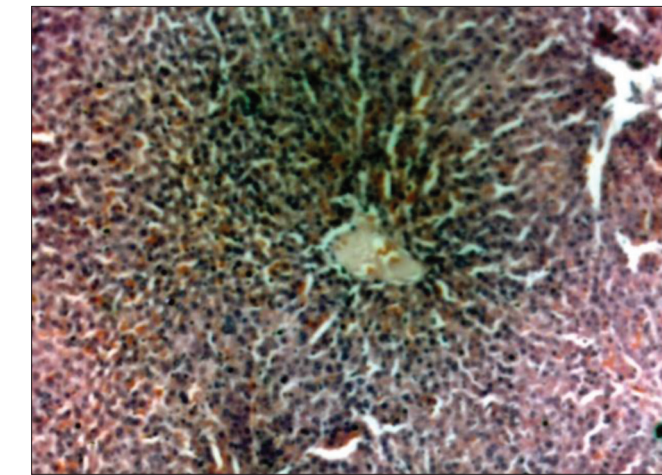
1.3 Group III: Partial-thickness mucosal necrosis



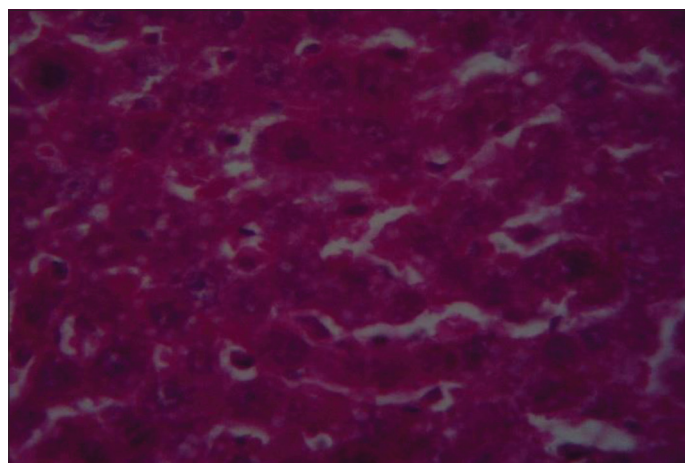
2.2B Group II: Microvesicular diffuse fatty change



2.2C Group II: Periportal inflammation

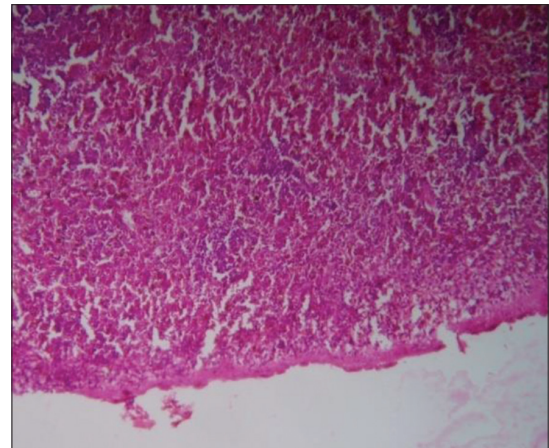


2.3A Group III: Mild congestion

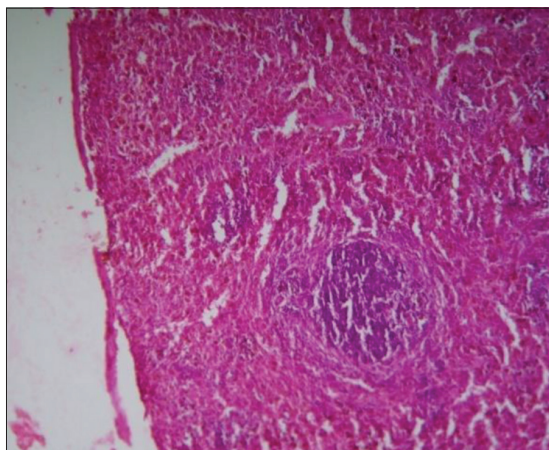


2.3B Group III: Regenerative hepatocytes

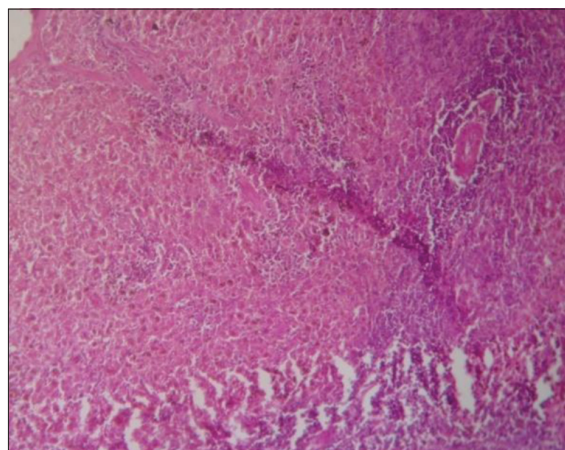
Plate 3 - Experimental animal study: Histopathology of spleen



3.1 Group I: Normal spleen



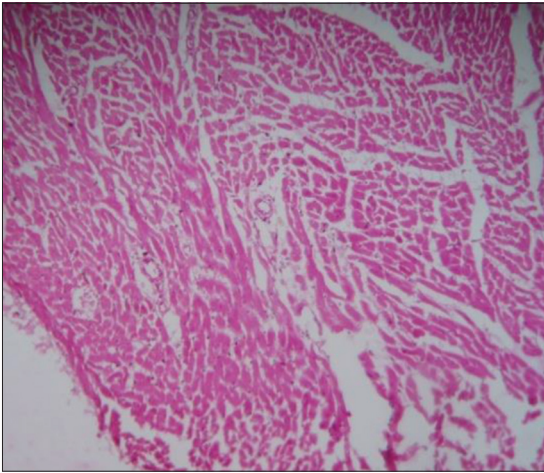
3.2 Group II: Sinusoidal congestion



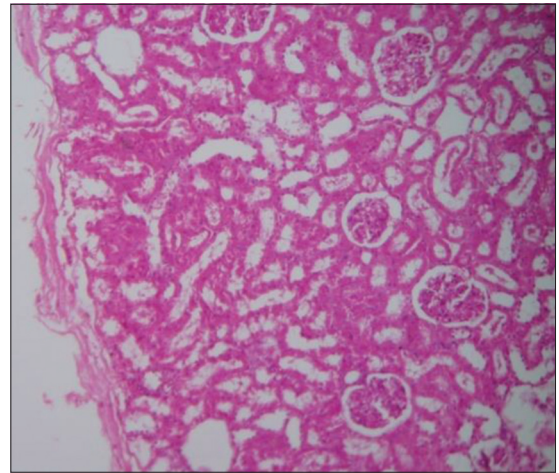
3.3 Group III: Sinusoidal congestion

Plate 4 - Experimental animal study: Histopathology of heart

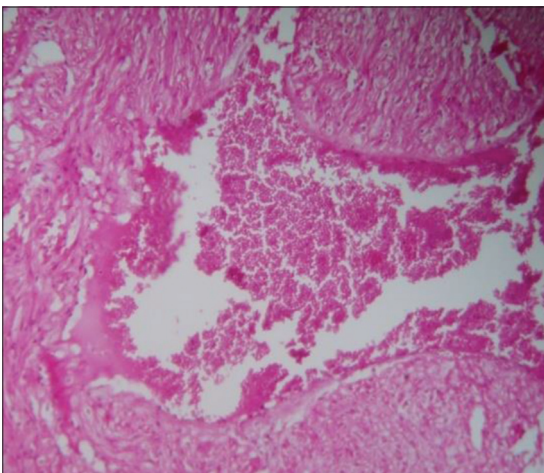
Plate 5 - Experimental animal study: Histopathology of kidney



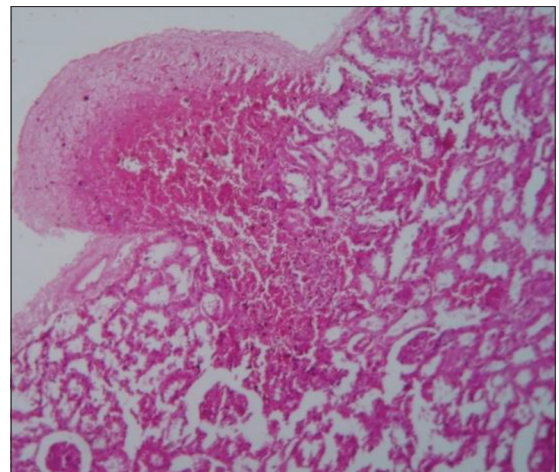
4.1 Group I: Normal heart



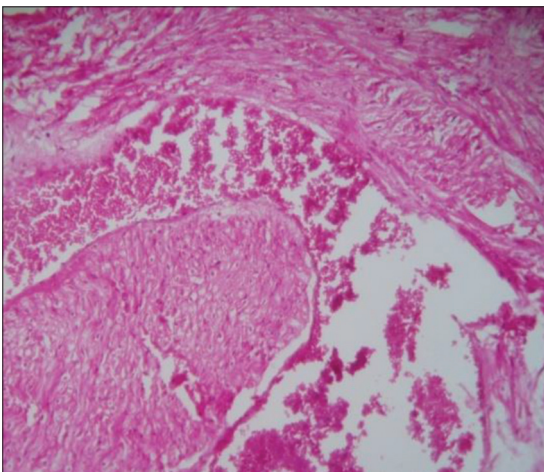
5.1 Group I: Normal kidney



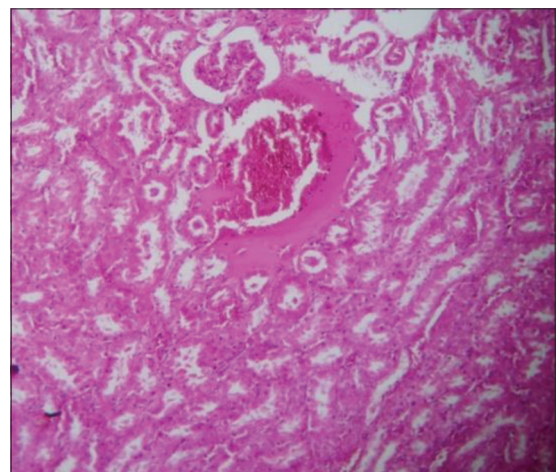
4.2 Group II: Hemorrhage



5.2 Group II: Capsular hemorrhage

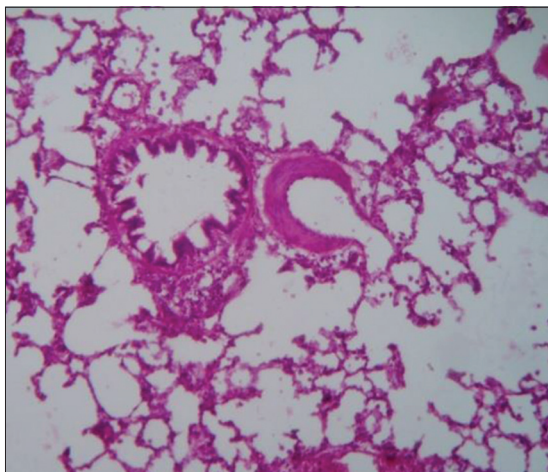


4.3 Group III: Hemorrhage

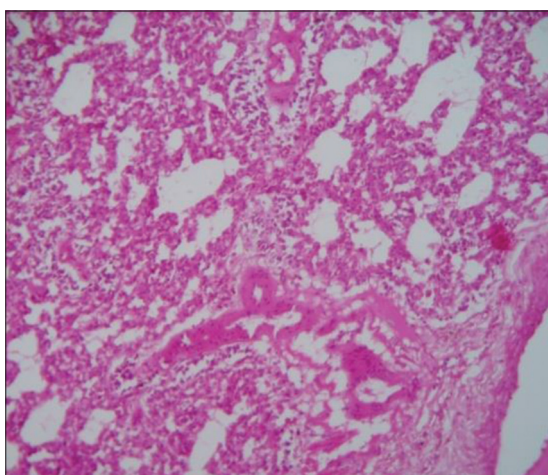


5.3 Group III: Parenchymal hemorrhage

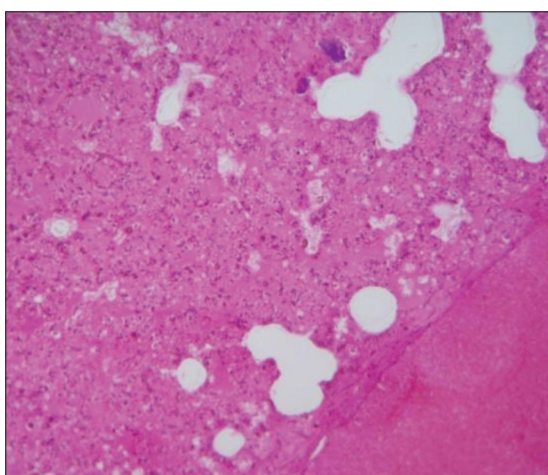
Plate 6 - Experimental animal study: Histopathology of lung



6.1 Group I: Normal lung



6.2 Group II: Inflammation and collapse



6.3 Group III: Severe collapse and edema

DISCUSSION

The main objectives of our experimental study were to study the toxicity profile of ABVD and COPP regimes in

albino rats and to assess the protective role of Vitamin E, an antioxidant, on drug-induced toxicities. Mean hemoglobin of Group II (group receiving anticancer drugs alone) was less compared to Group III (cytoprotective group), but P value was insignificant ($P = 0.8988$) (Table 7.1 and 7.2), indicating only minimal protection by Vitamin E for the cytoprotective groups. Regarding total count values, there was significant difference ($P = 0.0371, 0.021962$) between the groups (Table 8.1 and 8.2), indicating a protective role by Vitamin E for Group III. Platelet count did not show much variation in the three groups studied (Tables 9.1 and 9.2). Weight of animals at the 4th week was compared to the basal values. In the 4th week, both Groups II and III demonstrated weight loss; but the weight loss in Group III was significantly ($P = 0.0277, 0.026836$) (Table 10.1 and 10.2) much less, compared to Group II which again points toward the protection afforded by Vitamin E.

Several studies suggest that supplementation with antioxidants influences the response to chemotherapy as well as the development of adverse side effects related to chemotherapy.^[5-7] Administration of antineoplastic agents results in oxidative stress, that is, the production of free radicals and other reactive oxygen species (ROS). ROS cause or contribute to certain side effects that are common to many anticancer drugs, such as gastrointestinal toxicity and mutagenesis. ROS also contribute to side effects that occur only with individual agents, such as doxorubicin-induced cardiotoxicity and bleomycin-induced pulmonary fibrosis. Antioxidants detoxify ROS and can reduce or prevent many of these side effects and may enhance the anticancer effects of chemotherapy.^[8]

Histopathological Examination of the Internal Organs of Albino Rats Studied

Protective role of Vitamin E on liver and stomach in rats was studied by Canturk *et al.*^[9] The authors found that macroscopic and microscopic mucosal injuries were significantly greater in the control than the Vitamin E pre-treatment group. In our study, histopathological examination of the gastric mucosa revealed full-thickness surface epithelial necrosis in Group II (Plate 1.2) whereas Group III demonstrated only partial-thickness surface epithelial necrosis (Table 11; Plate 1.3). The necrosis could be due to the drug-induced changes in the gastric mucosa, and difference in severity in necrosis could be due to Vitamin E protection in Group III.

It was found in our study that spleen, kidney, and lungs were not protected from the toxic effects of the drugs by Vitamin E, as evidenced by the results (Tables 13, 15 and 16). In our study, the spleen showed sinusoidal congestion with hemorrhage when treated with cytotoxic chemotherapy irrespective of the use of the antioxidant Vitamin E (Plate 3). The kidney demonstrated tubular necrosis, subcapsular, and parenchymal hemorrhage in all groups except for the control (Table 15, Plate 5). The lungs showed evidence of inflammation and

edema in both Groups II and III. Some specimens showed collapse of a severe nature (Table 16, Plate 6).

A study was conducted by Legha *et al.*^[10] to investigate the effects of Vitamin E on Adriamycin cardiotoxicity. The occurrence of congestive heart failure in three patients and the observation of significant pathologic changes in endomyocardial biopsies in approximately half of the patients treated with a median cumulative Adriamycin dose level of 550 mg/m² indicate that Vitamin E does not offer substantial protection against Adriamycin-induced cardiac toxicity. In our study, the heart was also, not found to be protected by Vitamin E from these effects (Table 14, Plate 4).

In our study, histopathological examination of the liver showed marked toxic effects in Group II (Plate 2.2A, 2.2B and 2.2C), such as severe sinusoidal congestion, hemorrhage, hepatic necrosis, periportal inflammation, and diffuse fatty change. These changes were absent or minimal in Group III, with some specimens demonstrating regenerative hepatocytes (Table 12, Plate 2.3B). This further proves the scavenging role Vitamin E plays to counter the oxidative stress the liver is exposed to, during the course of these regimens.

The limitations of this study are that the data are representative alone and do not imply that all the animals in a particular group demonstrated that particular histomorphology.

CONCLUSIONS

The experimental animal study demonstrated a cytoprotective effect of Vitamin E against chemotherapy-induced organ toxicity in the animals studied. The protective effect was most evident in the stomach and liver and these observations appear promising. In the case of hematological toxicity, protection was only minimal. However, further experimental studies and extensive randomized controlled trials are warranted before implementing Vitamin E administration along with the ABVD/COPP regimen. The prime concern of chemotherapy is drug-associated oxidative stress, which results in many side effects. Use of antioxidants can be beneficial in this respect as they minimize the burden of free reactive radicals in cells and thus can decrease the duration of chemotherapy regimens.

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